

*Vector **NTI Advance***[™] **10**

Quick Start Guide

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Introduction

This Quick Start Guide is designed to get you started using Vector **NTI Advance**™ 10. It provides brief descriptions of the Vector **NTI Advance**™ 10 graphical user interface, including **Vector NTI Explorer** and the **Molecule Viewer**, and step-by-step instructions for using the most common features and functions of the software. The topics covered include displaying molecules, designing PCR primers, aligning molecules, performing a restriction analysis, and assembling contigs.

This guide assumes that you have a working knowledge of basic Microsoft® Windows® features and functions (how to open and save files, how to use your mouse, and so on) and that Vector **NTI Advance**™ 10 is installed on your computer.

Opening Vector NTI Advance™ 10

To launch Vector **NTI Advance**™ 10.

1. On your Windows® desktop, click on the **Start** button.
2. Select Programs | Invitrogen | Vector NTI Advance 10 to open the menu of Vector NTI Advance molecules.
3. Select **Vector NTI Explorer** to open the **Explorer** or **Vector NTI** to open the **Molecule Viewer**, as shown in Figure 1.

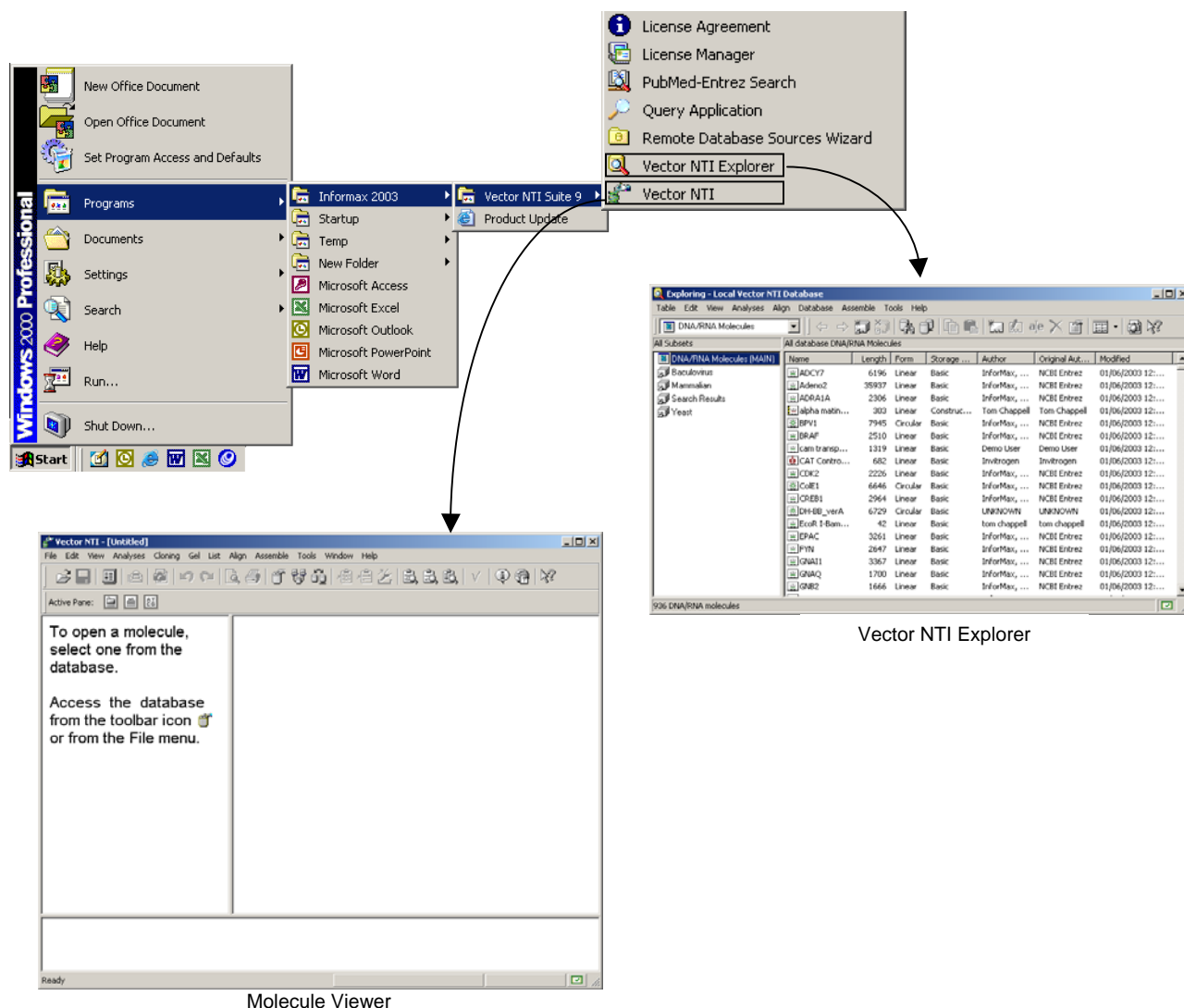


Figure 1. Opening the Molecule Viewer and Vector NTI Explorer.


You can configure the software to open both the **Molecule Viewer** and **Vector NTI Explorer** when you select **Vector NTI** from the **Start** menu.

1. In the **Molecule Viewer** window, go to the **Edit** menu and select **Options**.
2. In the **General** tab of the dialog, select the **Open Local Explorer At Startup** checkbox.
3. Click **OK** to make the change.

Vector NTI Explorer

Vector NTI Explorer is the main tool for accessing the information in your local **Vector NTI Advance™** database. Using the **Explorer**, you can import, open, export, and organize molecules and other database items, and launch other **Vector NTI Advance™** modules (Figure 2).

To launch **Vector NTI Explorer**:

- In the **Molecule Viewer**, click on the **Local Database icon** () , or
- From the Windows® Start menu, select Programs | Invitrogen | Vector NTI Advance 10 | Vector NTI Explorer
- A database in **Vector NTI Advance™** contains records for different types of molecular biology objects. Each database record includes all the information for that object (e.g., a DNA molecule record includes the DNA sequence, defined features of the molecule, and other information). Objects in the database can include molecules, analysis results, BLAST search results, citations, and other types of information.

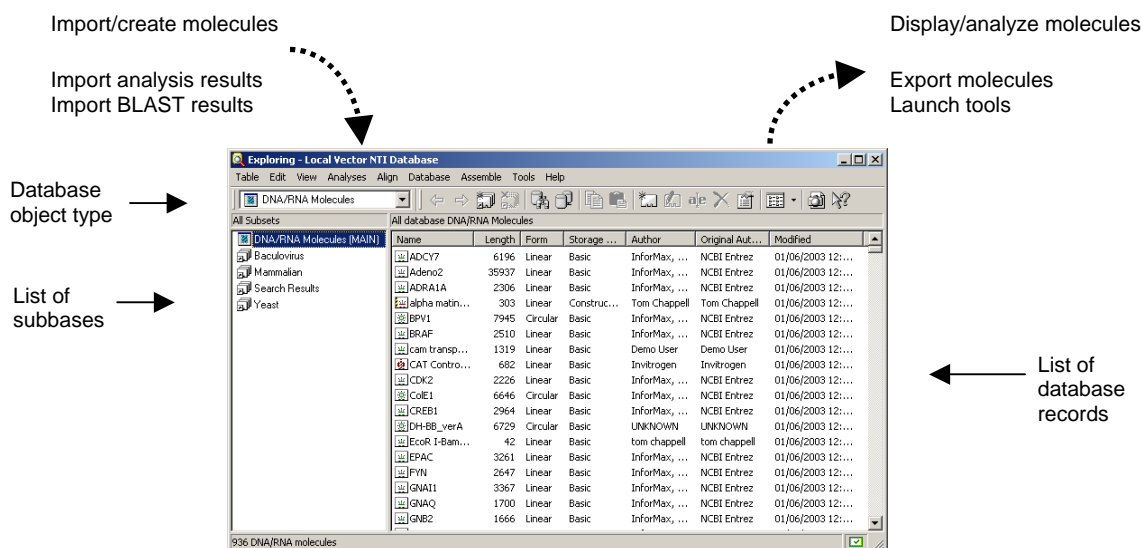


Figure 2. Vector NTI Explorer window.

Database objects in **Vector NTI Advance™** are categorized by type (DNA molecules, protein molecules, and so on). Some molecules are installed with the software. When you first open the software, **DNA/RNA Molecules** is the selected object type. Click on the dropdown list in the upper left corner of the **Vector NTI Explorer** to select from the other available database object types (Figure 3).

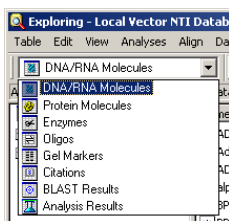


Figure 3. List of object types.

Continued on the following page

Vector NTI Explorer, continued

To open an object from the local database, double-click on the object name in the right-hand pane of the **Explorer**. Depending on the object type, information about that object may be displayed in a dialog box, or the object may be loaded into a viewer. For example, DNA, RNA, and protein molecules are displayed in the **Molecule Viewer**.



Note

When you install *Vector NTI Advance*[™], the default local database is created in a folder called **VNTI Database** in the root directory of your computer (e.g., C:\VNTI Database).


You can configure the software to open **Vector NTI Explorer** with the **Molecule Viewer** when you select **Vector NTI** from the **Start** menu.

1. In the **Molecule Viewer** window, go to the **Edit** menu and select **Options**.
2. In the **General** tab of the dialog, select the **Open Local Explorer At Startup** checkbox.
3. Click **OK** to make the change.

Molecule Viewer

The **Molecule Viewer** displays information about DNA, RNA, and protein molecules. To launch the **Molecule Viewer**:

- From the Windows® Start menu, select Programs | Invitrogen | Vector NTI Advance 10 | Vector NTI explorer, or
- Double-click on a molecule name in the **Vector NTI Explorer**.

To open a molecule from within the **Molecule Viewer**, click on the **Open button** () on the main toolbar and select the molecule name from the dialog box.

The molecule will be loaded into the **Molecule Viewer**.

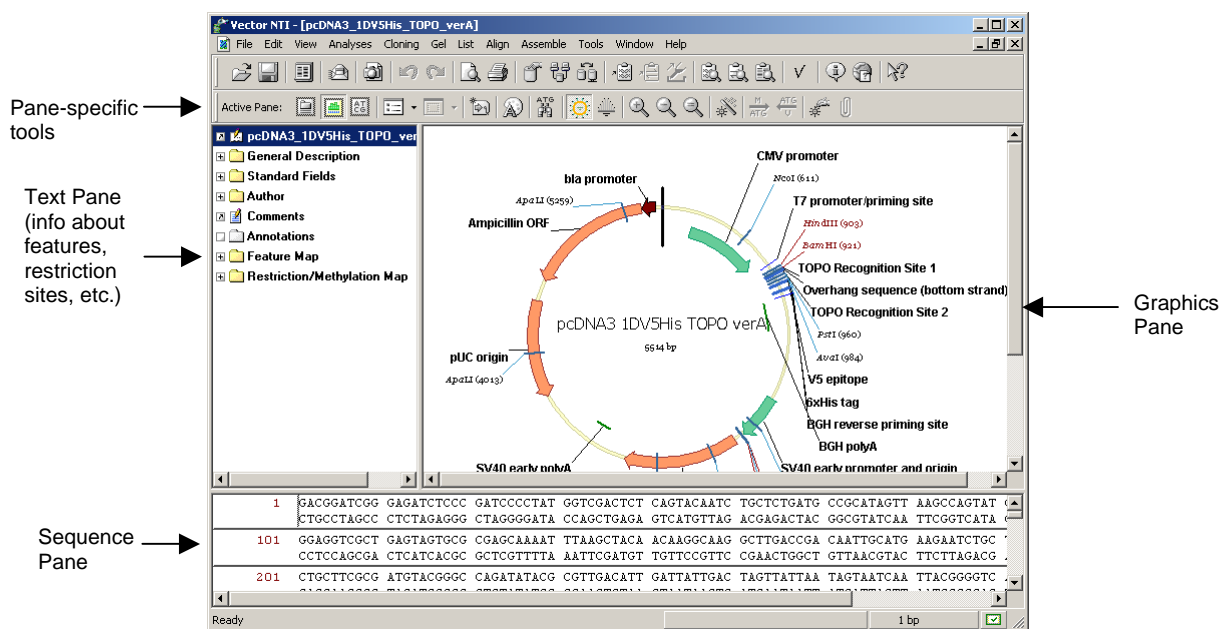


Figure 4. Molecule Viewer window for a DNA molecule.


The **Molecule Viewer** window has different panes for displaying different types of information about the molecule, as shown in Figure 4. Click inside a pane to make it the active pane. The available tools and right-click menu options will change depending on which pane is active.

Use tools on the dropdown menus and toolbars to add information about the molecule and perform various analysis functions, as described in the step-by-step instructions on the following pages.

Multiple molecules can be displayed separate windows of the **Molecule Viewer**.

Selecting and Editing Molecule Sequences

In the **Molecule Viewer**, you can select part of a molecule sequence in several different ways:

- Hold down the mouse button and drag the cursor across the sequence in the Sequence Pane or Graphics Pane (Figure 6).
- Go to the *Edit menu*, select **Set Selection**, and enter the sequence base-pair range in the dialog box.
- Click on a defined feature in the Graphics Pane.
- Click on a defined feature in the Text Pane, and click on **Find** () on the main toolbar.

The selected sequence will appear highlighted in both the Graphics Pane and the Sequence Pane.

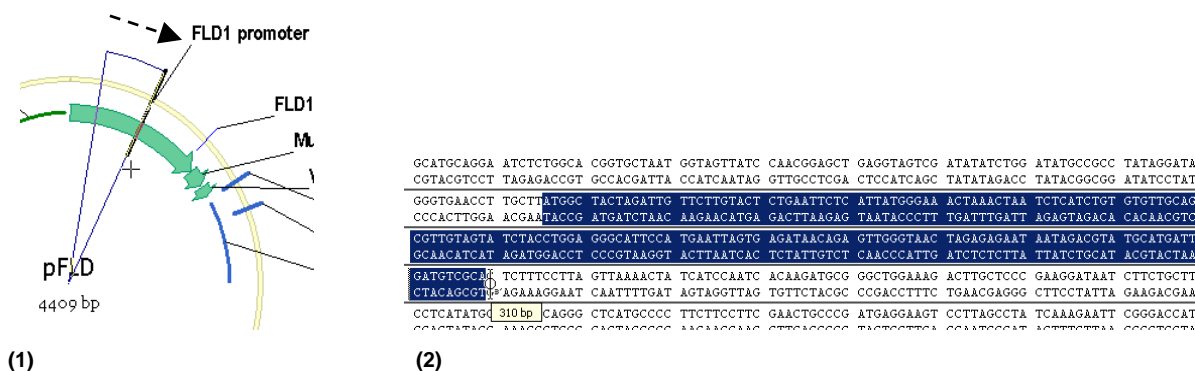


Figure 5. Selecting a DNA sequence by (1) dragging in Graphics Pane or (2) dragging in Sequence Pane.

To **copy** a molecule sequence:

1. Select it as described above.
2. To copy it to the Windows® clipboard, use the CTRL + C keyboard command, or

To copy the sequence as a text file, go to the *Edit menu* and select **Copy to > File**. You will be prompted to select a format and enter a name for the file.

To **delete** a molecule sequence:

1. Select it as described above.
2. Click on the DELETE key on your keyboard.

To **paste** a molecule sequence:

1. With the sequence in text format on the Windows® clipboard, click on the point in the Sequence Pane where you want to add the insert.
2. Click on CTRL + V on your keyboard.
3. The **Insert Sequence dialog** will open, displaying the sequence to be inserted.
4. Click on **OK** to complete the insertion.

Designing PCR Primers from a Sequence

Vector **NTI Advance**™ 10 can analyze a selected sequence and design PCR primers for it, based on parameters such as desired melting temperature (T_m), GC content, and amplicon length.

With a DNA or RNA molecule open in the **Molecule Viewer**:

1. Select the part of the sequence for which you want to design primers, as described on the previous page.
2. Go to the **Analyses menu** and select **Primer Design**.
3. Select **Find PCR Primers** to find primers within the sequence (Figure 6), or
Select **Amplify Selection** to find primers in the regions before and after the sequence (other amplification selections are available; see the Vector **NTI Advance**™ 10 User's Manual for more information).
4. In the dialog box, select the desired primer-design parameters. Note that most of these parameters have default values based on typical PCR primers.
5. Click on **OK**. The results will appear under **PCR Analysis** in the Text Pane.

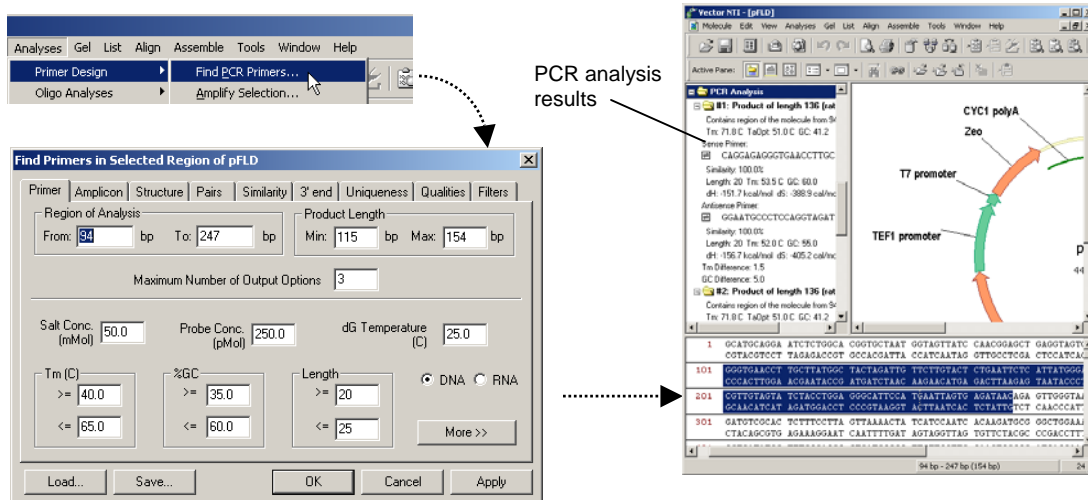


Figure 6. Designing PCR primers within a selected region.

To save the PCR analysis results as a separate object in the database:

1. Right-click on the **PCR Analysis** folder in the Text Pane.
2. Select **Save as Analysis Result**.

The saved results will be listed under the Analysis Results object type in the **Vector NTI Explorer** (Figure 7).

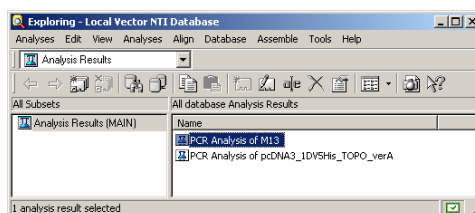


Figure 7. PCR analysis results listed in Vector NTI Explorer.

Identifying Open Reading Frames (ORFs)

Vector **NTI Advance**™ 10 can analyze a DNA/RNA molecule and identify the open reading frames (ORFs) in it, based on start and stop codons within the molecule.

With a DNA or RNA molecule open in the **Molecule Viewer**:

1. Go to the *Analyses menu* and select **ORF** (Figure 8).
2. In the dialog box, select the parameters for identifying and marking ORFs in the molecule.
3. When you click on **OK**, the sequences identified as ORFs will be marked with directional arrows in the Graphics Pane and Sequence Pane, and the ORFs will be listed in the Text Pane.
4. To identify an ORF in the different panes:
 - Click on a directional ORF arrow in the Graphics Pane to highlight its sequence in the Sequence Pane, or
 - Open a folder under **Open Reading Frames** in the Text Pane, right-click on the ORF name, and select **Find ORF** to highlight it in the Graphics and Sequence Panes.
5. To save an ORF to the feature map of the molecule, right-click on the ORF arrow in the Graphics Pane or the ORF folder in the Text Pane, and select **Add ORF to FMap**.

The figure illustrates the process of identifying Open Reading Frames (ORFs) in Vector NTI Advance 10. It shows the 'Analyses' menu with 'ORF...' selected, leading to the 'ORF Setup' dialog box. The dialog box contains settings for Minimum ORF Size (50 codons), Nested ORF (unchecked), Start Codons (ATG GTG), Stop Codons (TAA TGA TAG), and options for Incomplete ORFs and ORF Names. The 'ORF Setup' dialog box is shown with the 'OK' button highlighted. Below the dialog box, the 'Open Reading Frames' list in the Text pane is shown, listing various ORFs with their start and stop codons and regions. The 'pFLD' molecule viewer is shown with ORFs marked in the Graphics pane and highlighted in the Sequence pane. The 'pFLD' molecule viewer shows a circular map of the molecule with ORFs marked by green arrows. The 'Sequence pane' shows the DNA sequence with ORFs highlighted in blue. The 'Open Reading Frames' list in the Text pane shows the following ORFs:

- Open Reading Frames**
 - Direct strand**
 - Phase #1 (2 frames)**
 - 346(d1) (65 codons)**
 - Start codon: atg Stop codon: taa
 - Region: 346 - 540
 - 3895(d1) (84 codons)**
 - Start codon: glg Stop codon: tag
 - Region: 3895 - 4146
 - Phase #2 (2 frames)**
 - 3131(d2) (72 codons)**
 - Start codon: glg Stop codon: tag
 - Region: 3131 - 3346
 - 3698(d2) (124 codons)**
 - Start codon: atg Stop codon: tga
 - Region: 3698 - 4069
 - Phase #3 (2 frames)**
 - 1545(d3) (286 codons)**
 - Start codon: atg Stop codon: taa
 - Region: 1545 - 2402

The 'pFLD' molecule viewer shows a circular map of the molecule with ORFs marked by green arrows. The 'Sequence pane' shows the DNA sequence with ORFs highlighted in blue. The 'Open Reading Frames' list in the Text pane shows the following ORFs:

- Open Reading Frames**
 - Direct strand**
 - Phase #1 (2 frames)**
 - 346(d1) (65 codons)**
 - Start codon: atg Stop codon: taa
 - Region: 346 - 540
 - 3895(d1) (84 codons)**
 - Start codon: glg Stop codon: tag
 - Region: 3895 - 4146
 - Phase #2 (2 frames)**
 - 3131(d2) (72 codons)**
 - Start codon: glg Stop codon: tag
 - Region: 3131 - 3346
 - 3698(d2) (124 codons)**
 - Start codon: atg Stop codon: tga
 - Region: 3698 - 4069
 - Phase #3 (2 frames)**
 - 1545(d3) (286 codons)**
 - Start codon: atg Stop codon: taa
 - Region: 1545 - 2402

Figure 8. Identifying ORFs

Creating a Restriction Map

Vector **NTI Advance™ 10** can analyze a DNA/RNA molecule and identify the restriction sites in it, using the software's comprehensive library of restriction enzymes.

With a DNA or RNA molecule open in the **Molecule Viewer**:

1. Go to the **Analyses** menu and select **Restriction Analyses > Restriction Sites** (Figure 9).
2. In the **Restriction Map Setup** dialog, review the list of restriction enzymes in the **Use Enzymes:** field. These are the enzymes that will be used to identify the restriction sites. Click on the **< Add**, **> Remove**, and **>> Remove All** buttons to add and remove enzymes from the list.

Note: If you click on **< Add**, the **Choose Database Enzymes** dialog will open, listing all the enzymes in the database. Select enzymes in the list by clicking on them or click on the **Select All** button, and then click on the **OK** button to add them to the **Restriction Map Setup** dialog.

3. Click on **OK** in the **Restriction Map Setup** dialog. The restriction enzymes and their binding sites will be shown in the Graphics Pane and Sequence Pane. The specific cut site of each enzyme will be listed under **Restriction/Methylation Map** in the Text Pane.

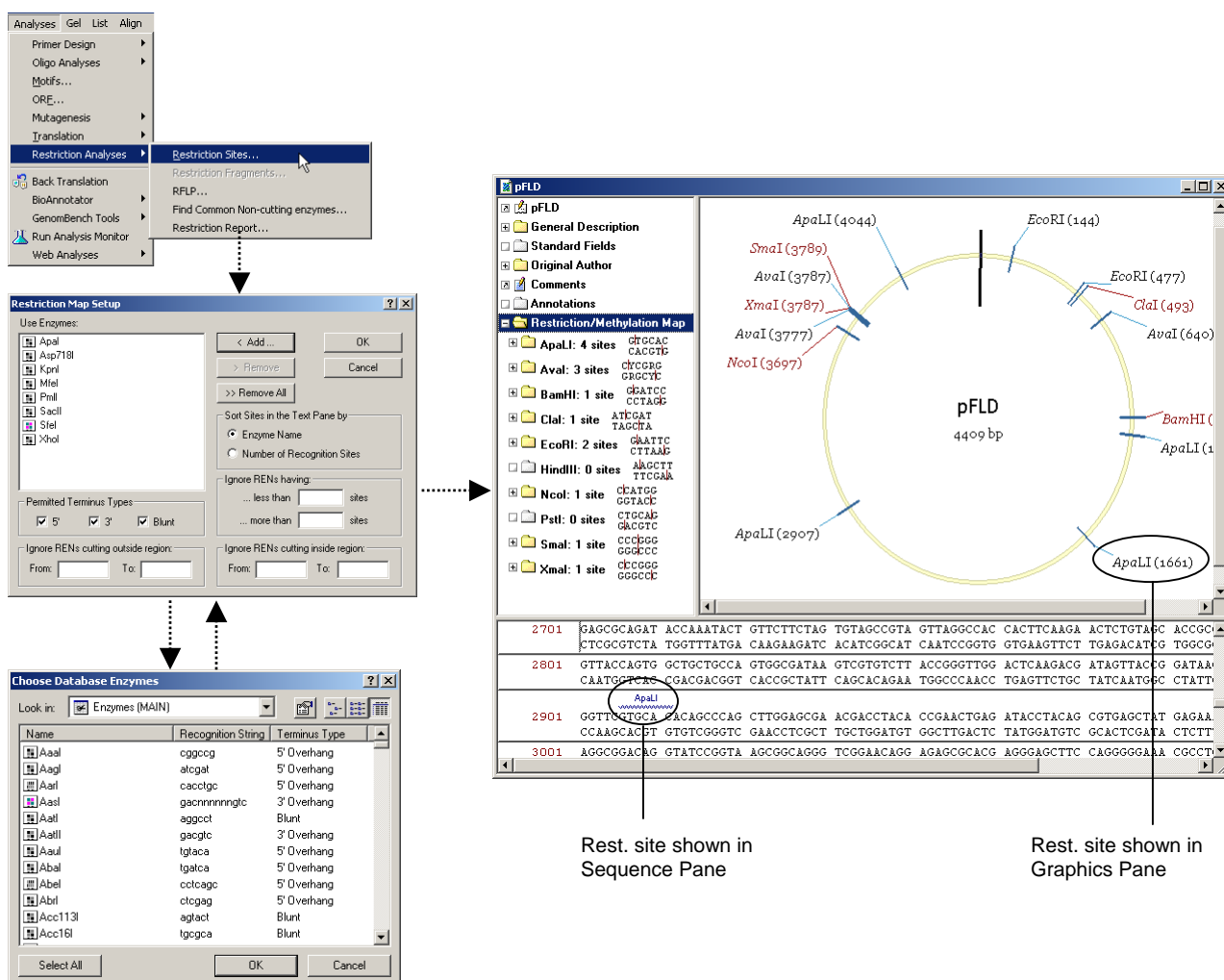



Figure 9. Creating a Restriction Map.

Aligning Molecules

Vector **NTI Advance**™ 10 can align the sequences of two or more DNA/RNA molecules. The tool for doing this is called **AlignX**. This tool can be launched from either the **Molecule Viewer** or **Vector NTI Explorer**.

To align sequences using **Vector NTI Explorer**:

1. In the **Explorer**, select the molecules that you want to align using SHIFT + CLICK or CTRL + CLICK key commands (Figure 10).
2. Go the *Align menu* and select **AlignX—Align Selected Molecules**. The **AlignX Window** will open, with the molecules you selected listed in the upper left Text Pane.
3. In the **AlignX Window**, use SHIFT + CLICK or CTRL + CLICK key commands to select two or more molecules in the Text Pane list to align.
4. To begin the alignment, click on the **Align button** () on the toolbar. The alignment may take several minutes, depending on the length and number of the molecules selected.
5. When the alignment is complete, the results are displayed in the **AlignX Window**, as shown in Figure 10. The **AlignX Window** has panes showing different similarity graphs and the points at which the sequences align.

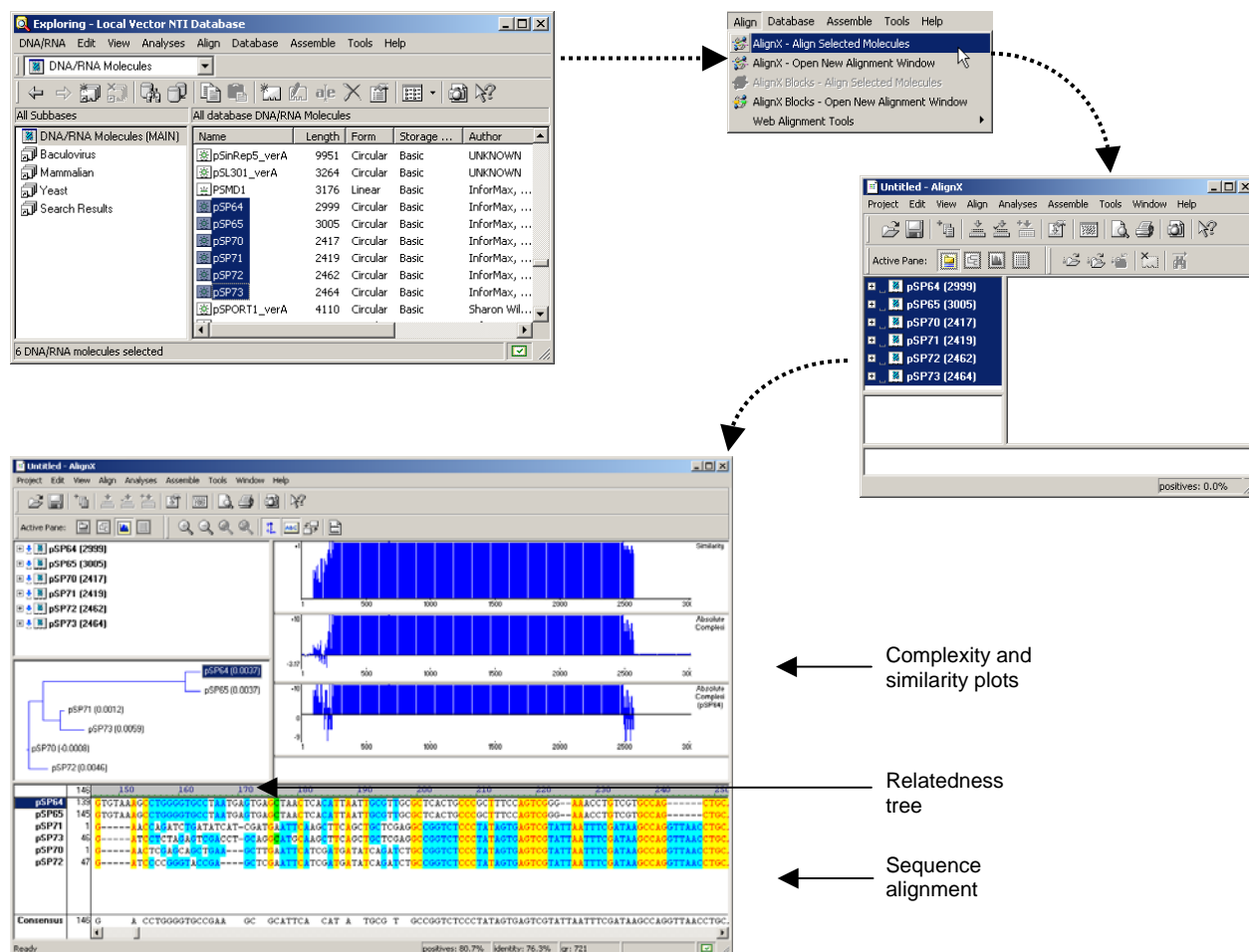



Figure 10. Aligning molecules.

Contig Assembly

Vector **NTI Advance™ 10** can be used to assemble DNA fragments—both text sequences and chromatograms from automated sequencers—into longer contiguous sequences or “contigs.” The tool for doing this is called **ContigExpress**.

In **Vector NTI Explorer** or the **Molecule Viewer**:

1. Go to the *Assemble menu* and select **ContigExpress—Open New Assembly Project** (Figure 11).
2. In the **ContigExpress Project Explorer**, go to the *Project menu* and select **Add Fragments >**. Select your fragment file type from the submenu list. The **Import Sequence dialog** will open.
3. In the **Import Sequence dialog**, navigate to the directory containing your fragment sequence files. Select the files and click on **Open**.
4. Depending on the file type, you may be prompted to list the fragments by their Windows® file names or by their internal fragment names. Select the desired option. The fragments will be loaded in the **ContigExpress Project Explorer**.
5. To view a particular fragment, double-click on it in the **Project Explorer** list. It will be loaded into the **Fragment Viewer**.
6. When you are ready to perform contig assembly, select the fragments in the **ContigExpress Project Explorer**.
7. Click the **Assemble Selected Fragments icon** () on the main toolbar. Fragments will be analyzed and assembled into one or more contigs, which will be listed in the **Project Viewer** along with the fragments in each contig.
8. Double-click on a contig in the list. It will be displayed in the **Contig Viewer**. The Sequence Pane at the bottom shows the sequence of the assembly. The Graphics Pane on the right shows the orientations of the fragments in the assembly. The Text Pane on the left lists the fragments in the assembly.
9. There are three trimming options in **ContigExpress**. Fragments can be trimmed for ambiguities, Phred quality scores, and vector contamination. Refer to the *Vector NTI Advance™ 10 User's Manual* for details.

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Contig Assembly, continued

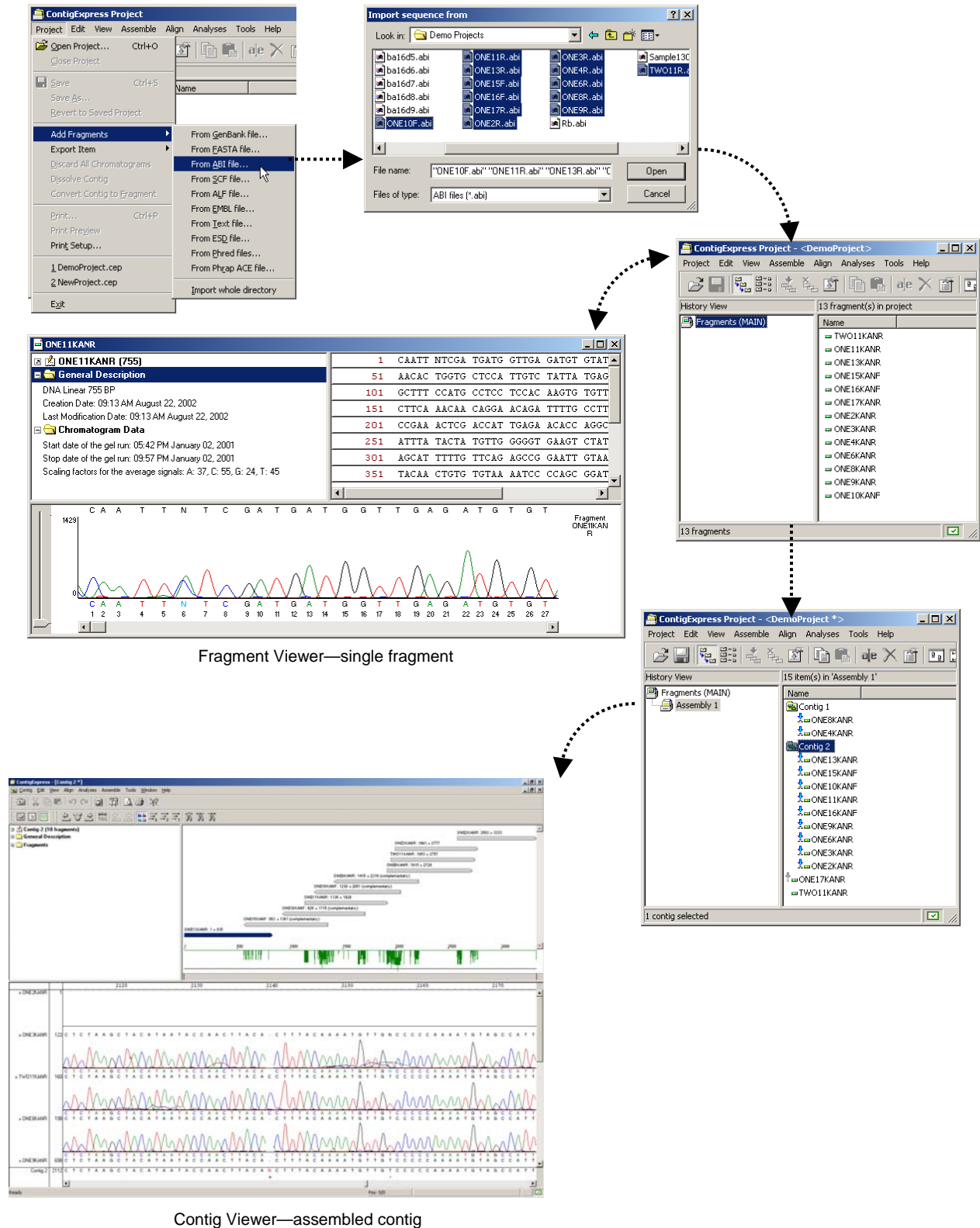


Figure 11. Assembling a Contig

Additional Information

Invitrogen's free technical support for Vector **NTI Advance 10** is available exclusively through the web. For more information, check out the eSupport section of the Vector NTI user community at <http://www.invitrogen.com/vectorNTIcommunity>.

To obtain personalized technical support by telephone or email, you must have an annual support contract. Academic and government users may purchase an Academic Vector NTI Support Contract through the User Community. Commercial users may purchase an Advanced Support Contract by contacting Invitrogen at bioinfosales@invitrogen.com (for the Americas) or eurobioinfoorders@invitrogen.com (for European/Middle Eastern/African/Asian Pacific customers).

For paid support, use the following contacts:

United States

Phone: 240-379-4240 or 877-357-3114 (Toll-free, U.S.)

E-mail: bioinfosupport@invitrogen.com

Europe, Middle East, Africa, Asian Pacific

Phone: +44 (0) 141 814 6350

Email: eurobioinfosupport@invitrogen.com

The Invitrogen Technical Support team may request that you run the Vector NTI System Information utility and email them the resulting file. Vector NTI System Information is a utility application, which gathers data about the system on which Vector NTI is installed and consolidates that information into one file. The system information file is helpful in the diagnosis and resolution of system-related Vector NTI problems.

